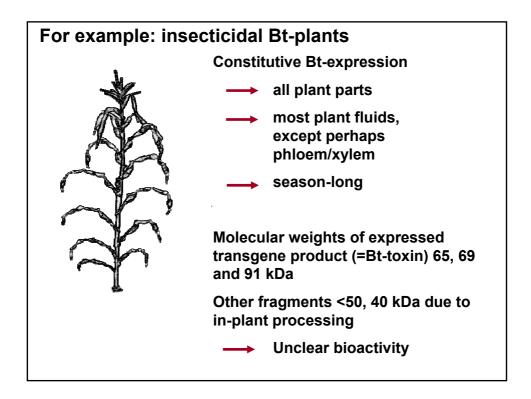


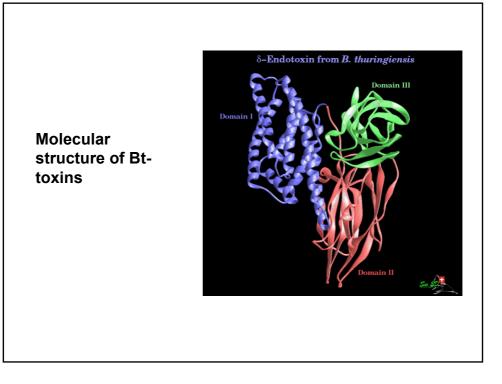


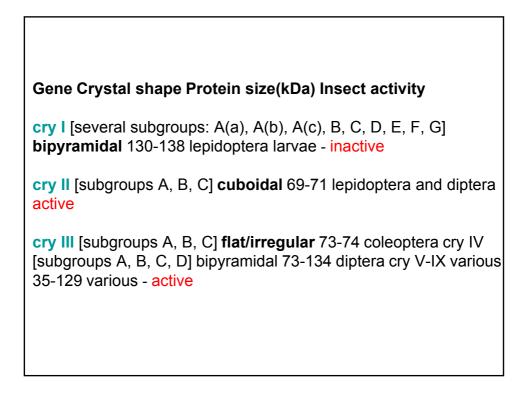
<u>Bt-Maize</u> produces one or two (or more?) toxin from a bacteria (*Bacillus thuringiensis* = Bt) that kill the LARVAE of a number of caterpillar and beetle species (Cry1, Cry2 and Cry3)











Some commercially available Bt varieties and target pests:

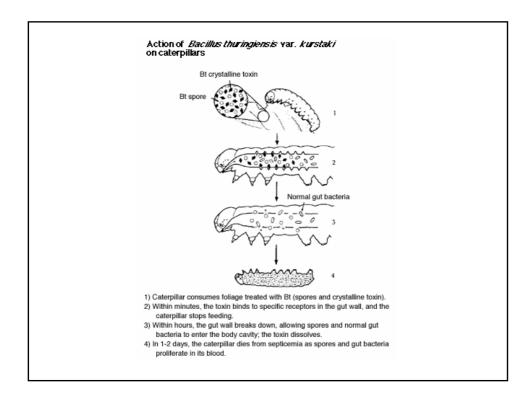
•Bacillus thuringiensis

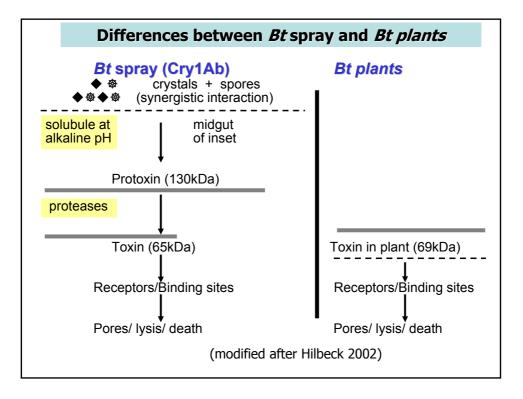
•var. *tenebrionis* - Colorado potato beetle and elm leaf beetle larvae

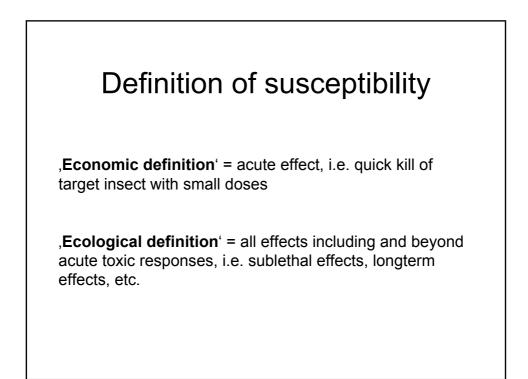
•var. kurstaki - caterpillars

•var. *israelensis* - mosquito, black fly, and fungus gnat larvae

•var. *aizawai* - wax moth larvae and various caterpillars, especially the diamondback moth caterpillar







New results on Bt mode of action

Broderick et al. 2006 PNAS September

Midgut bacteria required for Bt insecticidal activity

If target insects are treated with antibiotics Bt toxins do NOT work! Unclear why but apparently certain (which?) bacteria are necessary to induce mode of action.

Might explain all kinds of ,strange' non-target effects but would require concerted efforts in research.

Went astoundingly unnoticed by Bt-scientists

Reported non-target effects

Review by Hilbeck & Schmidt. 2006. Another view on Bt proteins – How specific are they and what else might they do? Biopesticides International – download at <u>www.gmo-guidelines.info</u>

In 27 (50%) of reviewed 54 studies reported negative effects on one or more of the tested parameters.

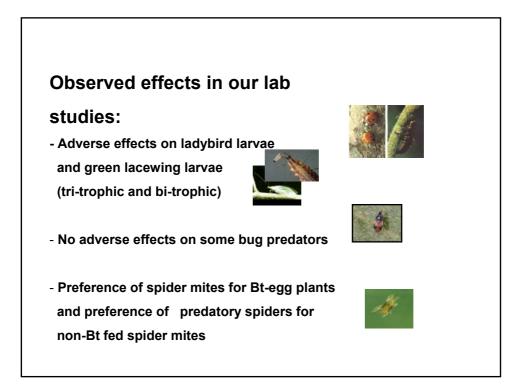
Positive effects were rare

The observed effects were in terms of degree and type of impact often unpredictable.

Reported non-target effects

"The mode of action of Cry1 toxins in non-target Lepidotera is presumed to be similar to that in target Lepidoptera. However, additional studies seem to be necessary to confirm this, in particular for non-target lepidoptera that exhibited only sublethal effects. Most notably, Deml et al. (1999) who conducted an extensive study with native Bt toxins found that also the coleopteranactive Cry3A toxins can have adverse effects on nontarget Lepidoptera.

Similarly, Hussein et al. (2005) and Hussein et al. (2006) reported deleterious effects on polyphagous moth *Spodoptera littoralis* when caterpillars were fed Cry3A-expressing potato foliage."



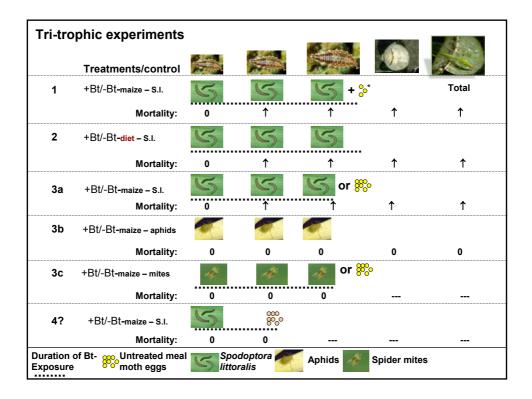


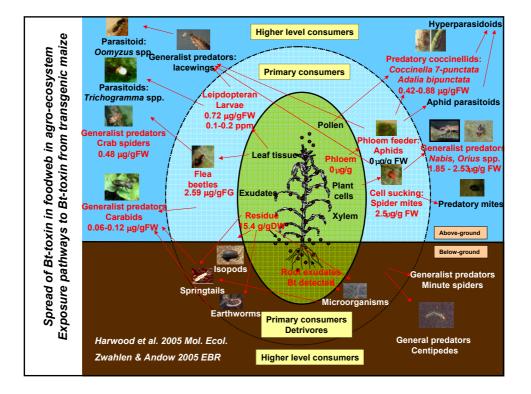
Feeding habit: Inject enzyms in prey, liquefied prey contents are sucked and ingested.

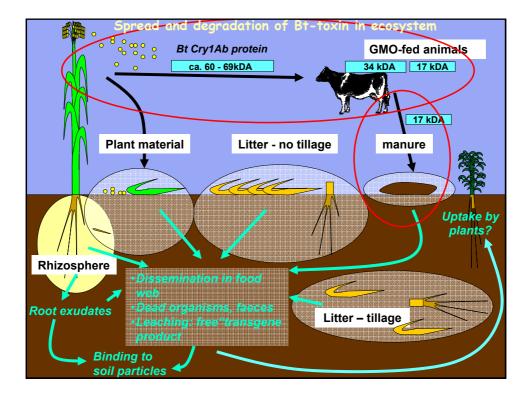
Prey: Larvae eat many other insects incl. fellow chrysopids. Preference for aphids if present. Optimal prey are small lepidoptera eggs.

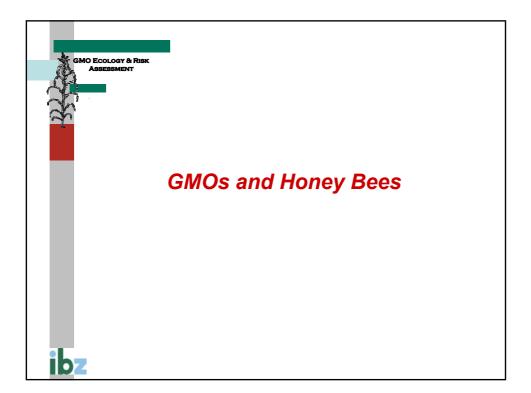
Ecotox testing approach: 'Bi-trophic' feeding studies

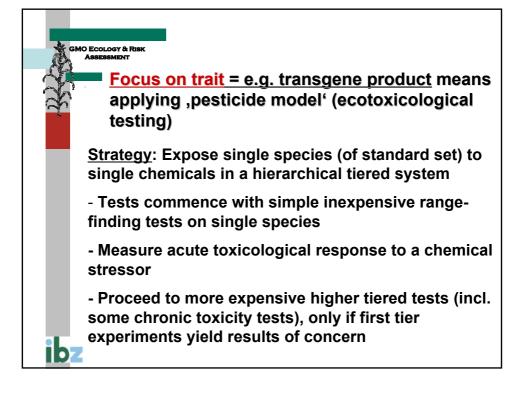
Bi-tro	phic experiments					
	Treatments/control			A COLOR	2 george	
1a	+Bt/-Bt – continual development	880	8800	88%		Total
	Mortality:	=	1	1	↑	↑
1b	+Bt/-Bt – continual development	800	800	8000		
	Mortality:	=	1	1	1	1
1c	Negative control	888	8800	<mark>880</mark> 0		
	Mortality:	=	=	=	=	=
2a	+Bt/-Bt – arrested development Time to death:	=	All test insects	die 		
2b	+Bt/-Bt – arrested dev. Followd by recovery Mortality:	* +8 ⁸ 8	800 0 =	800 =		
3	+Bt/-Bt	~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~	Exp	osure still needs	to be demonstra	ated
Duration Exposure	00-0	ving 👷		al Water dr w/wo Bt	ops co toxin ^{co} eg	ated meal moth gs w/wo Bt toxin











Standard non-target organisms tested according ,pesticide paradigm'

Water fleas (*Daphnia magna*) – <u>acute</u>, 48 hrs static renewal with <u>pollen</u> Springtail (*Folsomia candida*) – <u>chronic</u>, 28 days, <u>yeast</u> + test material Earthworm (*Eisenia foetida*) – 14 days, <u>soil</u> + test material Honey bee (*Apis mellifera*) – <u>acute</u>, 45 minutes, <u>undigested pollen</u> + water

Predatory/parasitoids insects

Hippodamia convergens - <u>adults</u> tested, <u>bitrophic</u> *Nasonia vitripennis* – <u>adults</u> tested; <u>pupal parasitoid of house flies</u>,

minor ecological relevance, <u>bitrophic</u> *Chrysoperla carnea* – larvae, bitrophic, coated meal-moth eggs, ca. 1 week

Testmaterial used:

- Lyophilized leaf protein as dietary test material

- Microbially produced, activated Bt-toxin

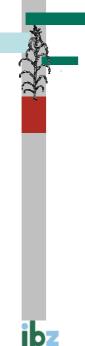
Test duration:

<u>Test endpoints:</u> toxicological parameters

- short time, acute

Testorganisms	Test method	Duration	OECD	
			Guideline No	
Water fleas (<i>Daphnia</i> spp.)	Acute immobilization Acute toxicity	24 - 96 hours	202	
Fish spp. (e.g. rainbow trout)	Acute toxicity	24 - 96 hours	203	
Fish spp.	toxicity of juvenile life stages	4 - 12 weeks	210	
Compost worm (<i>Eisenia</i> <i>foetida</i>)	Acute toxicity	7 – 14 days	207	
and mallards duck	Acute toxicity	14 - 21 days (few days treatment)	205	
Honey bees	Acute toxicity (oral) Acute toxicity	4 - 24 hours	New (1998) 213	

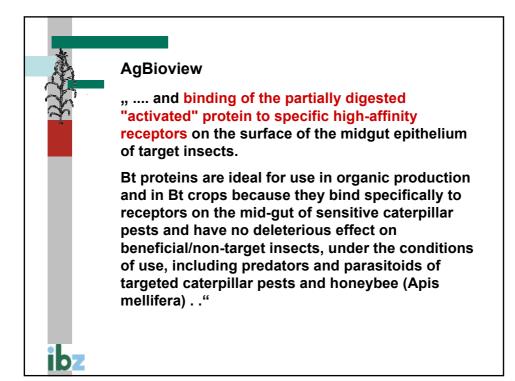
This is not sufficient! GM plants and their novel transgene products resemble plants rather than chemicals! ,Scientifically sound' testing must account for that!

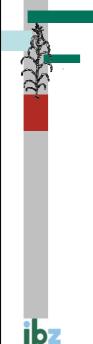


AgBioview

"There is extensive information on the lack of nontarget effects to diverse groups of beneficial insects including honey bees and other pollinators from Bt microbial preparations that contain Bt proteins.

--Bt proteins are ideal for use in organic production and in Bt crops because they are extremely selective and are toxic only to specific pests.... they bind specifically to receptors on the mid-gut of sensitive caterpillar pests and have no deleterious effect on beneficial/non-target insects under the conditions of use, including predators and parasitoids of targeted caterpillar pests and honeybees."

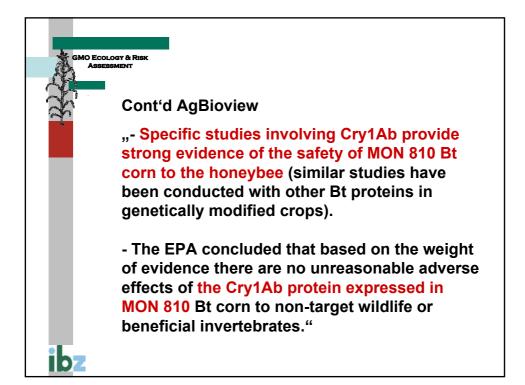




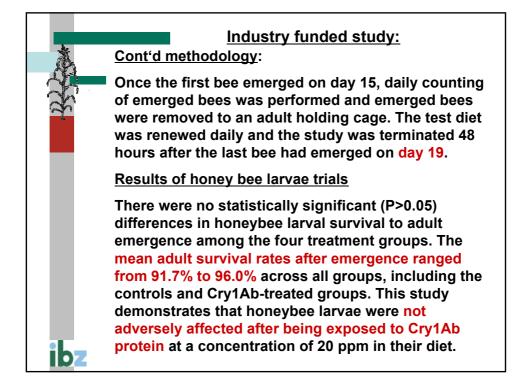
Cont'd AgBioview

"- Scientists perform extensive honeybee safety assessments on all insect-protected crops, including Bt corn and Bt cotton. The Bt proteins in these crops have been shown to have no adverse effect on the honeybee.

- EPA risk assessments have demonstrated that Bt proteins expressed in Bt crops do not exhibit detrimental effects to non-target organisms in populations exposed to the levels of Bt proteins produced in plant tissues."



Industry funded study: Methodology: Honey bee larvae were exposed to Cry1Ab protein in their natural diet by including a maximum hazard dose (20 parts per million in distilled water mixed with honey) in developing brood cells. This maximum nominal concentration of 20 ppm was approximately 100 times greater than the maximum expected Cry1Ab protein level in MON 810 pollen. In addition to this treatment group, a negative control group was treated with distilled water. Another control group was treated with heat-attenuated (inactivated) Cry1Ab protein (20 ppm), and one set of larvae received no treatment (untreated control). At least 50 bees (1 to 4 days old) were in each replicate, and there were three replicates for each group. The treatments were administered to each larval cell ibz through an electronic micro-applicator, which delivered 5 microliters of the test diet.



Industry funded study:

Methodology for adult bee trial:

.... mixing the appropriate amount of the insecticidally-active Cry1Ab protein with a honeywater (50-50) syrup to a concentration of 20 parts per million (microgram protein/g diet; ppm). The negative control group was fed the same diet with the exception that no Cry1Ab protein was added to the honey-water mixture. A second control group was fed heat-attenuated (inactivated) Cry1Ab protein at the same concentration (20 ppm) as the treatment group.

Results of adult experiment:

ibz

Adult honeybees exposed to the Cry1Ab protein in a honey-water solution for 9 days at a concentration of 20 ppm showed no signs of treatment-related mortality or toxicity. At the end of the testing period, the mortality percentage was calculated for each group. Mortality in the treatment and the negative control groups was 16.20% and 22.28%, respectively. The heatattenuated control group mortality was 32.59%. Mortality showed a sharp increase in all three groups from days 6 through 9. At the termination of the test, the highest mortality was observed in the group that was fed the heat-attenuated Cry1Ab protein diet, while the lowest mortality was observed in the group that was fed the Cry1Ab protein diet.

